

Sugar Beet Artificial Seeds an Overview

Wessam M. Rslan*

Agricultural Genetic Engineering Research Institute; Agricultural Research Center, Egypt.

*To whom correspondence should be addressed: wessam.rslan@ageri.sci.eg



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Abstract

Artificial seed propagation of crops broadens the horizon of plant biotechnology and farming. The technology offers techniques for micropropagulated seed analogs such as axillary leaves, embryogenic calli, somatic embryos, apical shoot tips, and protocorm-like organs. Micropropagules are embedded in gelling medium and carboxyl methyl cellulose active coatings. A variety of plant species, such as mulberry, sandalwood, cardamom, banana, sugar beet, maize, and relative, have recorded encapsulation of micro shoots and somatic embryos and subsequent recovery of full plantlets. This knowledge has shown that artificial seed manufacturing is possibly helpful for the propagation of economically significant species ' inferior hybrids on a big scale. Artificial seed development can only succeed with effective upstream manufacturing of micropropagules and downstream germination procedures for an elevated proportion of plant regeneration as one of the significant value-added plant tissue culture goods. Different micropropagules were regarded for the manufacturing of artificial seeds; however, mostly favored were somatic embryos and axillary stem buds. As micropropagules, somatic embryos were used to create artificial seeds in a wide range of fruit and plant organisms, which include *Daucus carota*, *Picea abies*, *Arachis hypogaea*, *Medicago sativa*, *Psidium guajava*, and *Vitis vinifera*. The review illustrated the concept of synthetic seeds and encapsulation procedure of sugar beet.

Keywords: Artificial seed, plant biotechnology, embryogenic, sugar beet, micropropagulated.

Introduction

Ara et al. (1) described artificial plants as synthetically encapsulated somatic embryos, shoot tips, axillary flowers or another meristematic tissue employed for seed sowing and capable of converting into whole plants under in vitro and in vivo circumstances and maintaining their potential even after storage. The first one to suggest and handle encapsulated somatic embryos was Murashige (2). He also noted to the transfer option and used it as natural plants. Reddy et al. (3) clarified that the cultivation of plants using artificial plants from somatic embryos or other vegetative propagules opens up new agricultural and forestry technologies. A variety of plant species synthetic seeds have been effectively researched in plant propagation.

Encapsulating agents

Kitto and Janick (4) discovered that the most appropriate medium for the encapsulation of somatic embryos was 'Polyox,' water-soluble resin. Redenbaugh et al. (5) suggested that magnesium alginate should be the most appropriate to encapsulate somatic embryos in alfalfa, celery, cauliflower and rose. Artificial seed performance relies on the spatial, qualitative, quantitative availability of growth regulators and nutrients together with an ideal physical setting (6). Mariani (7) indicated that gibberellic acid (GA3) and saccharose had an adverse impact on eggplant germination of synthetic seeds. In the meantime Refouvelet et al. (8) used BA (5 mg / l) + 1/2 MS + NAA (0.01 mg / l) to encapsulate *Syringa vulgaris*. Pattnaik and Chand (9) axillary buds, Murashige and Skoog (10) medium (MS) without hormones and MS + 6-benzyladenine (BA, 4.4 µM) have been used as artificial endosperm in *Morus* species.

Saiprasad (11) revealed that sodium alginate would be the most widely recognized hydro-gel and was regularly used as a matrix for artificial plants due to its small toxicity, low price, rapid gellation and bio-compatibility. Many researchers noted that the introduction to the encapsulation solution of aquatic cyanobacterial samples (12), bactericides and activated carbon (13), pesticides, fertilizers, microorganisms (Rhizobia), mycorrhiza fungi (14), fungicides (15) can safeguard encapsulated propagules from microorganisms, decrease the discharge of toxic compounds and improve the germination ability of seeds.

Encapsulation procedure

The hydro-gel encapsulation technique established by (5) was the most suitable technique for producing synthetic seeds. In this technique, by combining with calcium free liquid MS medium, sodium alginate of varying levels (2 to 5

percent) was formed and then the explants were blended with the solution. Together with the sodium alginate solution, explants were sucked with a pipette and dropped into calcium chloride pool in which the ion exchange reaction happens and sodium ions were substituted by calcium ions comprising alginate beads. It is necessary to complete the entire method under aseptic circumstances. The capsule size relies on the pipette nozzle's inner diameter. The beads structure and size relies on sodium alginate quantity, calcium chloride solutions, and complexation duration. Redenbaugh (16) proposed using a multi nozzle pipette in this embryo stream through the internal pipette and the solution of the alginate flows via the exterior pipette.

Artificial seeds germination and field planting

Several researchers (17–20) recorded successful field cultivation and transformation of natural plants. The synthetic plants in the future, particularly for the extremely requested species (21), may be an option planting material intended for the forestry sector. Artificial plants would enable plant propagules to be planted directly into the greenhouse or field, circumventing many of the additional phases (21). Fujii et al. (22) discovered that the maturation of ABA somatic alfalfa embryos yielded an elevated soil transformation rate of 48% to 64%. Adding fungicide to alginate beads avoids contamination and increases the sustainability of mulberry seeds in soil (23) when sown. Fujii et al. (24) revealed effective field planting with 23 percent crop transformation of alfalfa artificial plants obtained from calcium alginate embryoids.

Nieves et al. (25) revealed that artificial plant sugarcane crops were larger and had a lower diameter at eight months, but at 12 months these distinctions faded. No variations in all parameters assessed among both artificial seed-derived plants and plants based from the other two techniques (traditional and isolated plant techniques) were discovered with regard to sugar assessment and yield. Asmah et al. (21) and Ma et al. (26) recorded the efficient germination level in *Acacia* hybrid (73.3 to 100%) and *Pseudostellaria heterophylla* (80%). In *Podophyllum peltatum*, sugar beet and *Stevia rebaudiana*, Rizkalla et al. (27) and Nower (28) noted that crop development improved by adding mannitol and/or sorbitol to the medium.

Types of synthetic seeds

Two forms of synthetic seeds have been established, i.e. desiccated and hydrated synthetic plants, according to the current literature. The synthetic desiccated seeds were first launched either directly or encapsulated in polyox from somatic embryos, followed by their desiccation (29).

Desiccation has been accomplished either linearly through chambers of decreasing relative humidity slowly over a period of one or two weeks or swiftly by leaving the petri dishes overnight on the bench at the laminar airflow room (30). The hydrated artificial seed technology was first developed by encapsulating *Medicago sativa* (16) hydrated somatic embryos. These artificial hydrated seeds are employed to create plant species that are recalcitrant and susceptible to desiccation in their somatic embryos. Hydrated artificial seeds are usually ready in a hydrogel capsule by encapsulating somatic embryos or other propagules. Several techniques for producing hydrated artificial seeds were investigated, mostly using calcium alginate encapsulation (16).

The genetic stability of synthetic seeds

Artificial seeds were commonly used in many plant species for micro-propagation. Molecular researches have begun from the last decade to determine genetic stability of plantlets derived from synthetic seeds, but no changes have been revealed at the biochemical and/or molecular scales. Many studies (31) endorsed the prospective benefit of synthetic seeds for genetically identical to natural seeds. Gangopadhyay et al. (32) investigated the genetic structure of plantlets obtained from encapsulated *Ananas comosus* micro shoots using RAPD and ISSR technologies (33, 34). Bekheet (35) indicated that both plantlets obtained from encapsulated bulblets and usually in vitro were genetically comparable to those obtained from in vivo in *Allium sativum*.

Narula et al. (36) used RAPD assessment to explore in vitro plantlet genetic structure obtained from *Dioscorea bulbifera* encapsulated plant advice. Srivastava et al. (37) reported that the analysis of *Cineraria maritana's* RAPD patterns revealed a median ratio of resemblance of 0.944, confirming the molecular consistency of crops extracted from encapsulated micro-shoots followed by six months of storage. Tabassum et al. (38) investigated the genetic consistency of synthetic seeds obtained from mother crops and somatic embryos and discovered similarity in *Cucumis sativus* using RAPD markers.

Mishra et al. (39) also studies the genetic consistency of crops obtained from encapsulated microshoots in *Picrorhiza kurrooa* using RAPD profile cluster analysis. Lata et al. (40) used ISSR and gas chromatography (GC) study of six significant cannabinoids to examine the genetic structure of synthetic seed based crops of *Cannabis sativa* and demonstrated homogeneity in the regrown clones and the mother plant. Shoot tips are by far the most genetically consistent, but in callus and protoplast culture there is a strong probability of genetic shift (41–43).

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